

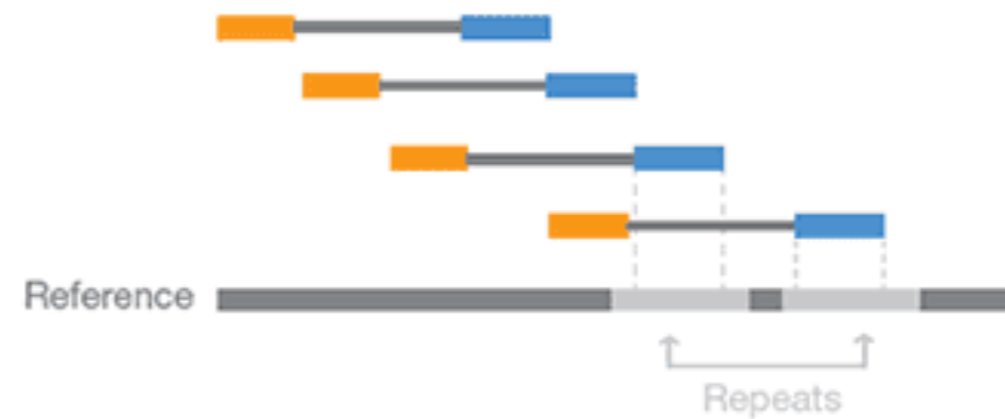
High-Throughput Sequencing: from Raw Reads to Variants

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Paired-End Reads



Alignment to the Reference Sequence



Paired-end sequencing enables both ends of the DNA fragment to be sequenced. Because the distance between each paired read is known, alignment algorithms can use this information to map the reads over repetitive regions more precisely. This results in much better alignment of the reads, especially across difficult-to-sequence, repetitive regions of the genome.

HTS data analysis is ultimately about placing reads and accounting for uncertainty

Sources of Error

- (Library Preparation)
- Sequencing
- Read Cleaning
- Assembly
- Mapping
- Post-mapping Processing (e.g., indel realignment, etc.)
- Variant Calling
- Post-processing



Goals

- Introduce a (general) workflow for data analysis
- Describe the data structures of common files
- Perform analyses using the Unix command line
- After: Tiago will (among other things) analyze the same dataset using Galaxy
- Please ask me about your own datasets!

Recommendations

If you are going to be analyzing large datasets or lots of libraries, using the command line or custom scripts may be the best way to go

- Unix (+ a shell language like bash)
- A scripting language (R or Python, possibly Perl)
- Application development: a compiled language (C, C++)

Lots of help available in several great communities



Exercises

- All commands in the text
- Starting from *cleaned reads*
 - You need to do some processing beforehand!
- We'll stop periodically and examine some of the files that we've been generating

Mouse exome capture example

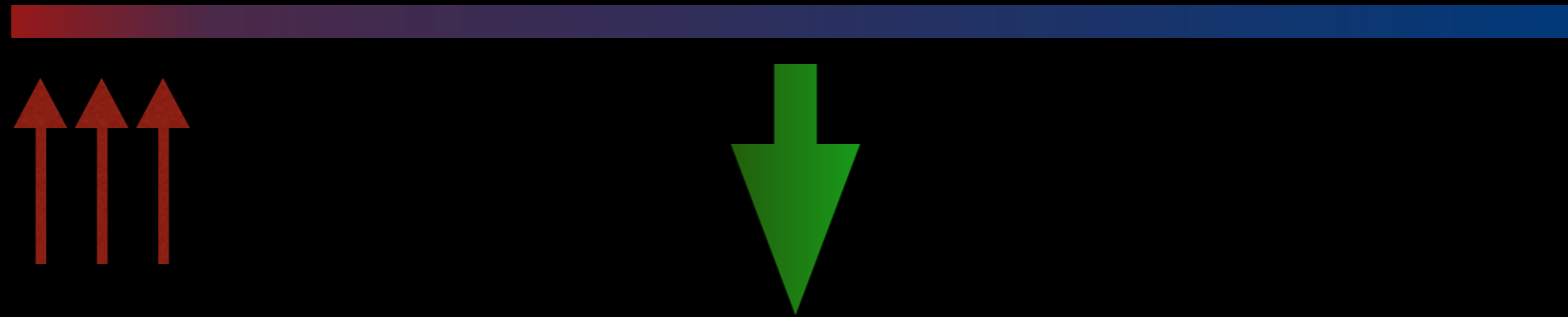
Mus spretus (Algerian mouse)

- 55 Mbp capture
 - Genome is ~2.8 Gbp (~2%)
- Carry over annotation information from the reference
- In place of the whole genome, we will be working with a 20 Mbp region of chromosome 1 containing approximately 50 transcripts

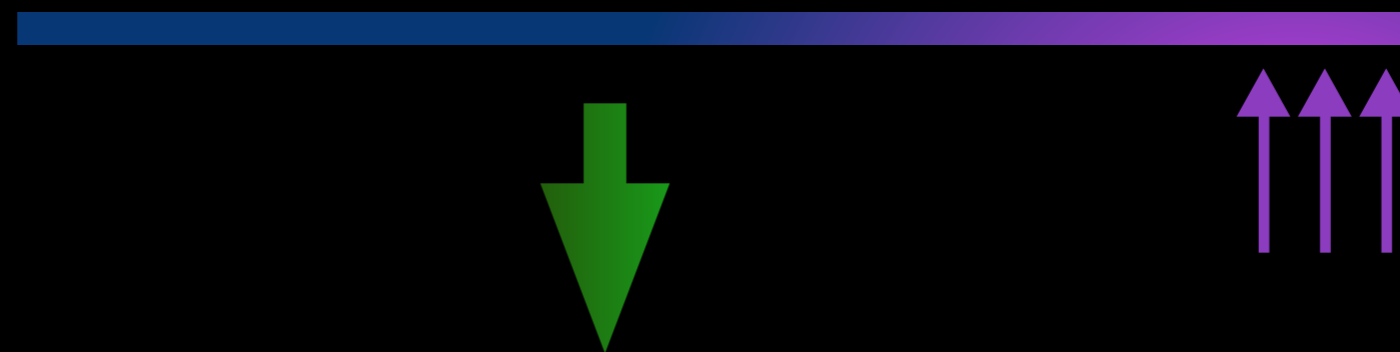


Pre-processing of Reads

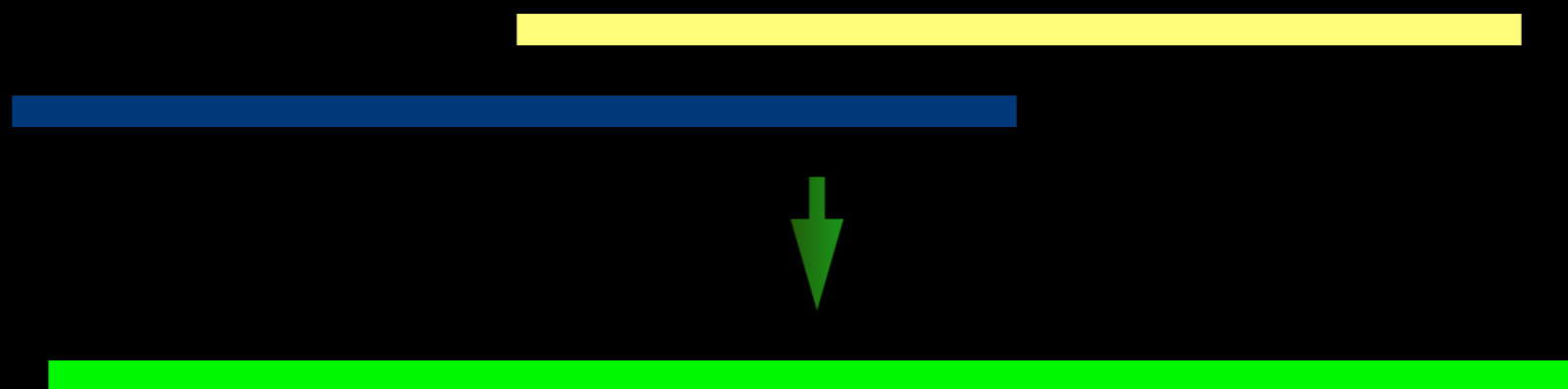
Removal of low-quality base calls



Removal of residual adapter sequence



Merge reads that overlap



The FASTQ format

```
congen@congen-VirtualBox:~/data/brice/test$ zcat 10252.final.fq.gz | head -n 12
@HS3:309:D2385ACXX:5:2210:7285:54270
TATCCAGCCAGCCTGGCTTAGATGGTGAGTGAGCGCCAGGCCAATGAGGAACCTGTGCCATGGACCGGGCCTAGTCAGCTCCCCTCAATTCGTGGGAATC
+
BBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBBB
@HWI-ST665R:136:C1G9MACXX:2:1302:14922:92414
TATCCAGCCAGCCTGGCTTAGATGGTGAGTGAGCGCCAGGCCAATGAGGAACCTGTGCCATGGACCGGGCCTAGTCAGCTCCCCTCAATTCGTGGGAATC
+
CCCCFFFFFHGHHHIIFIGIJJGIJ@ABFHGGHEBGAGHBHBFEGGHHGIJHIGGGHIJJIGEHB?CB>B@@BCCCCDDCACCBDDBDDDDDB>8<>?ABC
@DQNZZQ1:722:C2J9EACXX:7:2106:18279:13220
AAATGCCACGGACTGGGCTCAGTAGGCCCCCCTCAATCCATGGGAATCAGGGTTTCGGACAGATGGGCACAGAGTCGGTGAAAATAGGGTGACAAACAGACAGGACATAAGGAAGTGTGCTGAATCTGAATGT
+
<@@DDDDDEDADAFGIBBHHGFG@9CGBHIIHIEDHB)=BBCFHGGA?CDGEHB?ECE?=A>CD5DA<GCF@8F:?=HGGED?BCGE@HG?FBGEHEBDBEIJJIFHH>IIIIIGIIGGGEHHFHDDDDDB@@
```

Four lines per read:

1. Description - starts with @
 - @[Machine Identifier]:[Run Identifier]:[Flowcell ID]:[Flowcell Lane]:[Tile]:[x-coordinate]:[y-coordinate]
2. The base calls
3. +
4. Quality scores (ASCII)

The Variant Call Format

```
##reference=file:///home/congen/data/brice/test/chr1.20mbp.fa
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT 10252
chr1 608 . A G 34.74 . GT:AD:DP:GQ:PL ./.:1,4:5:6:62,6,0
chr1 622 . C T 14.68 LowQual . GT:AD:DP:GQ:PL ./.:0,8:8:3:41,3,0
chr1 657 . T C 14.68 LowQual . GT:AD:DP:GQ:PL ./.:3,5:8:3:41,3,0
chr1 659 . G A 14.68 LowQual . GT:AD:DP:GQ:PL ./.:6,1:7:3:41,3,0
chr1 1034 . T C 33.74 . GT:AD:DP:GQ:PL ./.:0,2:2:6:61,6,0
chr1 1820 . A G 15.65 LowQual . GT:AD:DP:GQ:PL ./.:0,1:1:3:42,3,0
chr1 3114 . A C 13.72 LowQual . GT:AD:DP:GQ:PL ./.:0,1:1:3:40,3,0
chr1 4332 . A C 11.83 LowQual . GT:AD:DP:GQ:PL ./.:0,5:5:3:38,3,0
chr1 5812 . C T 56.74 . GT:AD:DP:GQ:PL ./.:0,2:2:6:84,6,0
chr1 5837 . G A 56.74 . GT:AD:DP:GQ:PL ./.:0,2:2:6:84,6,0
chr1 5874 . C A 47.74 . GT:AD:DP:GQ:PL ./.:0,2:2:6:75,6,0
chr1 6024 . A G 11.83 LowQual . GT:AD:DP:GQ:PL ./.:0,1:1:3:38,3,0
chr1 6042 . T G 173.77 . GT:AD:DP:GQ:PL 0/1:28,8:36:99:202,0,981
chr1 6053 . G A 49.77 . GT:AD:DP:GQ:PL 0/1:33,5:38:78:78,0,1160
chr1 6062 . T C 1435.77 . GT:AD:DP:GQ:PL 1/1:0,37:37:99:1464,111,0
chr1 6064 . A G 1452.77 . GT:AD:DP:GQ:PL 1/1:0,37:37:99:1481,111,0
chr1 6065 . T A 1380.77 . GT:AD:DP:GQ:PL 1/1:0,37:37:99:1409,105,0
chr1 6073 . T G 1506.77 . GT:AD:DP:GQ:PL 1/1:0,38:39:99:1535,114,0
chr1 6075 . C A 1578.77 . GT:AD:DP:GQ:PL 1/1:0,40:40:99:1607,120,0
chr1 6111 . A G 1922.77 . GT:AD:DP:GQ:PL 1/1:1,50:51:99:1951,112,0
chr1 6156 . C A 188.77 . GT:AD:DP:GQ:PL 0/1:55,11:66:99:217,0,1988
chr1 6157 . A C,T 2271.77 . GT:AD:DP:GQ:PL 1/2:0,53,10:63:99:2300,347,188,1953,0,1923
chr1 6161 . T A 2384.77 . GT:AD:DP:GQ:PL 1/1:0,61:61:99:2413,184,0
chr1 6162 . G A 2286.77 . GT:AD:DP:GQ:PL 1/1:1,60:61:99:2315,142,0
chr1 6167 . A G 2497.77 . GT:AD:DP:GQ:PL 1/1:0,64:64:99:2526,193,0
chr1 6180 . C A 50.77 . GT:AD:DP:GQ:PL 0/1:55,7:64:79:79,0,1958
chr1 6186 . A G 1390.77 . GT:AD:DP:GQ:PL 0/1:16,40:58:99:1419,0,421
chr1 6187 . C T 44.77 . GT:AD:DP:GQ:PL 0/1:49,6:58:73:73,0,1740
chr1 6192 . T A,G 1801.77 . GT:AD:DP:GQ:PL 1/2:0,9,42:51:99:1830,1511,1483,319,0,193
chr1 6204 . C T 1479.77 . GT:AD:DP:GQ:PL 1/1:1,40:41:86:1508,86,0
chr1 6206 . A G 1351.77 . GT:AD:DP:GQ:PL 1/1:3,38:42:28:1380,28,0
chr1 6213 . G A 1126.81 . GT:AD:DP:GQ:PL 0/1:4,36:40:20:1155,0,20
chr1 6218 . G C 1199.77 . GT:AD:DP:GQ:PL 1/1:0,33:34:99:1228,99,0
chr1 6223 . G C 1146.77 . GT:AD:DP:GQ:PL 1/1:0,31:34:93:1157,93,0
chr1 6231 . G T 850.77 . GT:AD:DP:GQ:PL 1/1:0,24:24:72:879,72,0
```

GT: genotype

AD: allelic depth

DP: depth of coverage

GQ: genotype quality

PL: Phred-scaled genotype likelihoods